

# High-resolution multi-z confocal microscopy with a diffractive optical element: supplement

**BINGYING ZHAO,<sup>1,\*</sup>  MINORU KOYAMA,<sup>2</sup>  AND JEROME MERTZ<sup>3</sup> **

<sup>1</sup> *Department of Electrical and Computer Engineering, Boston University, 44 Cummington Mall, Boston, MA 02215, USA*

<sup>2</sup> *Department of Cell and Systems Biology, University of Toronto, 1265 Military Trail, Scarborough, ON M1C1A4, Canada*

<sup>3</sup> *Department of Biomedical Engineering, Boston University, 44 Cummington Mall, Boston, MA 02215, USA*

\**byzhao@bu.edu*

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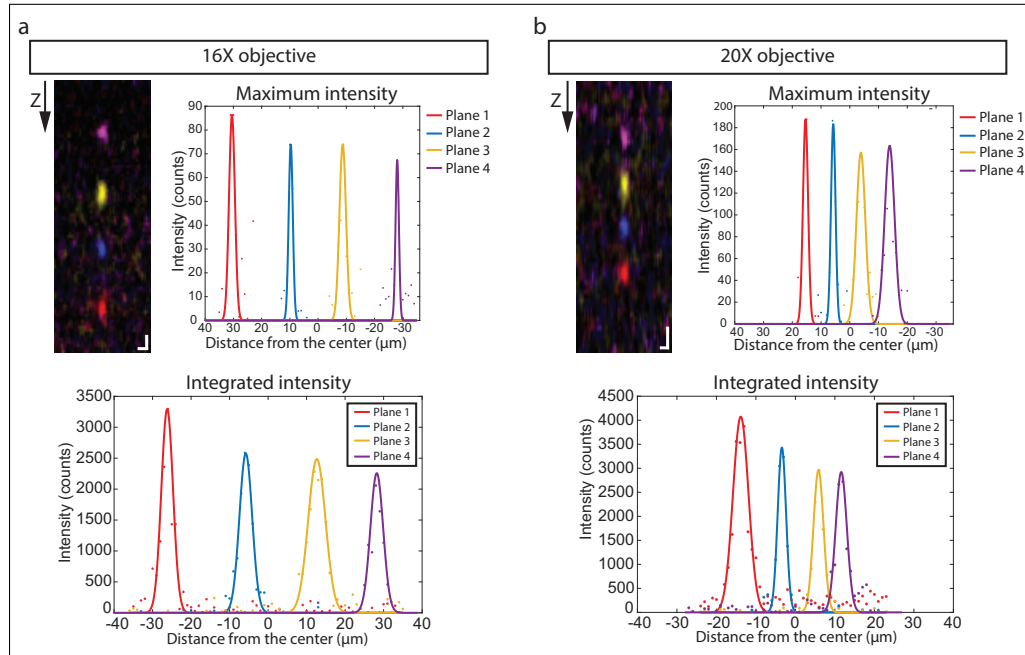
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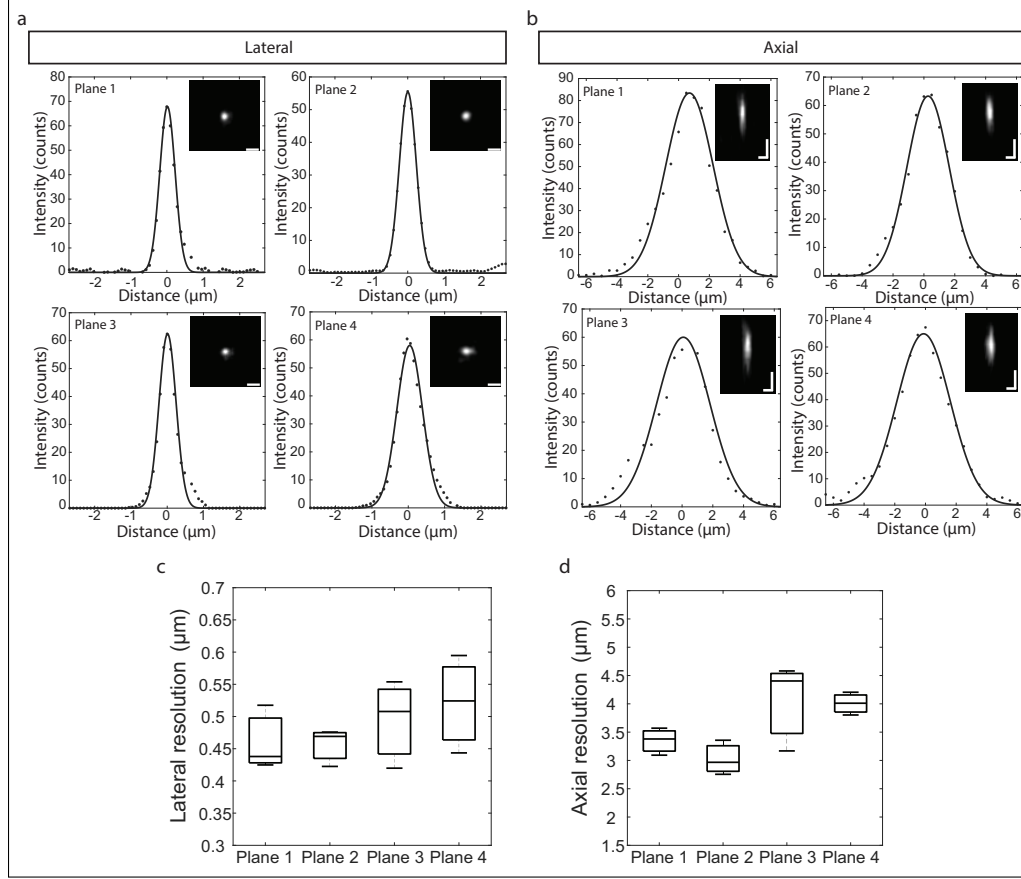
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# High-resolution multi-z confocal microscopy with a diffractive optical element

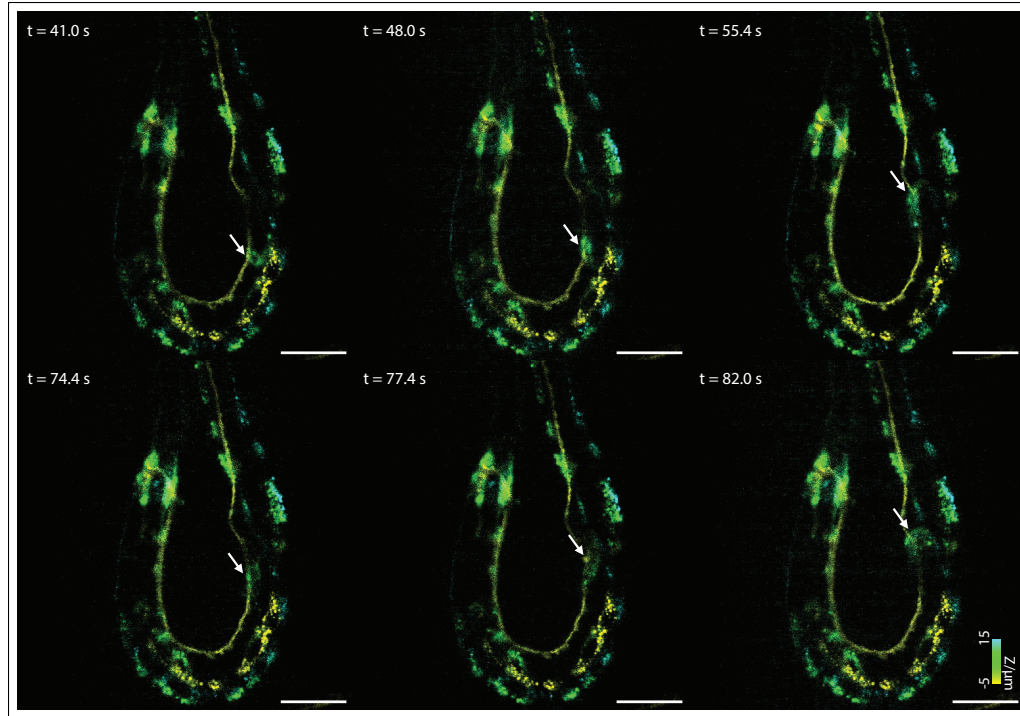
## 1. FIGURES



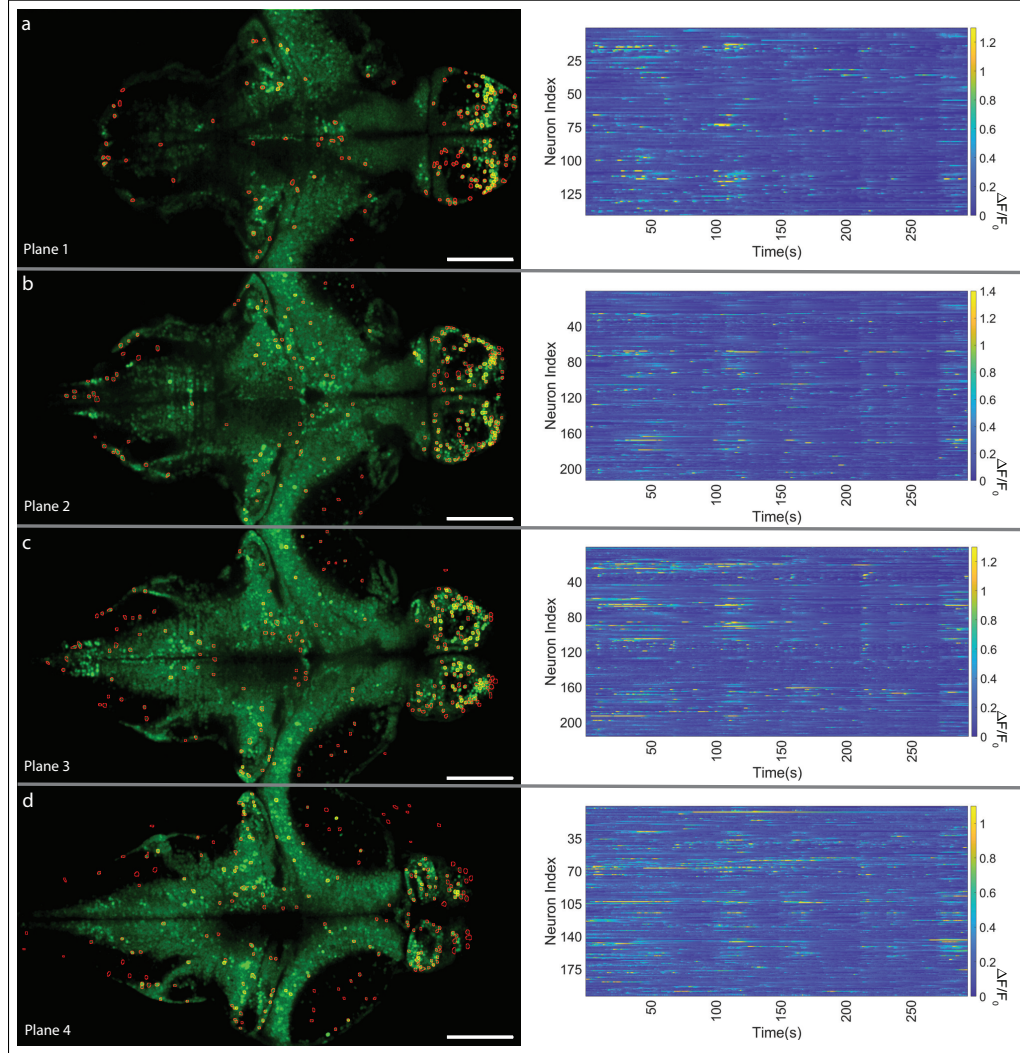
**Fig. S1.** Raw (un-normalized) intensity produced by a 0.2  $\mu\text{m}$  fluorescent bead when axially scanned through the imaging planes with (a) a 16 $\times$  objective ( $f_{obj} = 12.5\text{mm}$ ), and (b) a 20 $\times$  objective ( $f_{obj} = 9\text{mm}$ ). Maximum intensity: maximum intensity produced by a single fluorescent bead when axially scanned through the four imaging planes. Integrated intensity: intensity integrated over the x,y plane produced by a single fluorescent bead when axially scanned through the four imaging planes. Average post-objective laser power: 1 mW (for both objectives). Horizontal scale bar: 1  $\mu\text{m}$ . Vertical scale bar: 5  $\mu\text{m}$ . Planes 1-4: deepest to shallowest.



**Fig. S2.** Lateral (a) and axial (b) intensity produced by a 0.2  $\mu\text{m}$  fluorescent bead through the four planes. Planes 1-4: deepest to shallowest. Horizontal scale bar: 1  $\mu\text{m}$ . Vertical scale bar: 5  $\mu\text{m}$ . Lateral (c) and axial (d) resolution as measured with 0.2  $\mu\text{m}$  fluorescent bead ( $n = 12$ ). Centerline, medians; limits, 75% and 25%; whiskers, maximum and minimum. Planes 1-4: deepest to shallowest.



**Fig. S3.** *In vivo* calcium imaging of *C. elegans* at different times with color corresponding to depth. Scale bar:  $50\text{ }\mu\text{m}$



**Fig. S4.** *In vivo* calcium imaging of zebrafish brain. Planes 1-4: deepest to shallowest. Left: max-min projection from separate planes. Right: activity of identified neurons from separate planes. Scale bar: 100  $\mu\text{m}$

## 2. TABLE

**Table S1. Description of optical components used**

Component	Description	Part number, Vendor
L1	Plano-Convex spherical lens, $f = 25\text{ mm}$ , $\phi = 1\text{ in}$	LA1951-A-ML, Thorlabs
L2	Plano-Convex spherical lens, $f = 250\text{ mm}$ , $\phi = 1\text{ in}$	LA1461-A-ML, Thorlabs
L3	Plano-Convex spherical lens, $f = 40\text{ mm}$ , $\phi = 1\text{ in}$	LA1422-A-ML, Thorlabs
L4	Plano-Convex spherical lens, $f = 30\text{ mm}$ , $\phi = 1\text{ in}$	LA1805-A-ML, Thorlabs
L5	Plano-Convex spherical lens, $f = 125\text{ mm}$ , $\phi = 1\text{ in}$	LA1986-A-ML, Thorlabs
L6, L8	Achromatic doublets, $f = 100\text{ mm}$ , $\phi = 1\text{ in}$	AC254-100-A-ML, Thorlabs
L7	Achromatic doublets, $f = 300\text{ mm}$ , $\phi = 2\text{ in}$	ACT508-300-A-ML, Thorlabs
L9	Achromatic doublets, $f = 250\text{ mm}$ , $\phi = 1\text{ in}$	AC254-250-A-ML, Thorlabs
SL	Scan lens, $EFL = 70\text{ mm}$	CLS-SL, Thorlabs
DM	Dual-edge laser dichroic beamsplitter, $25.2\text{ mm} \times 35.6\text{ mm}$	Di01-R488/561-25x36, Semrock
GR	Galvo-resonant scanners, $8\text{ kHz}$	LSK-GR08, Thorlabs
DOE	Multifocal diffractive optics element, $\phi = 9.2\text{ mm}$	MF-005-R-Y-A, Holo/or
RP1 - 4	$150\text{ }\mu\text{m}$ rounded aperture, $\phi = 1\text{ in}$	214-0688, National Aperture
D1 - 4	Silicon photomultipliers, pixel pitch= $50\text{ }\mu\text{m}$ , $\phi = 1.5\text{ mm}$	S14420-1550MG, Hamamatsu